

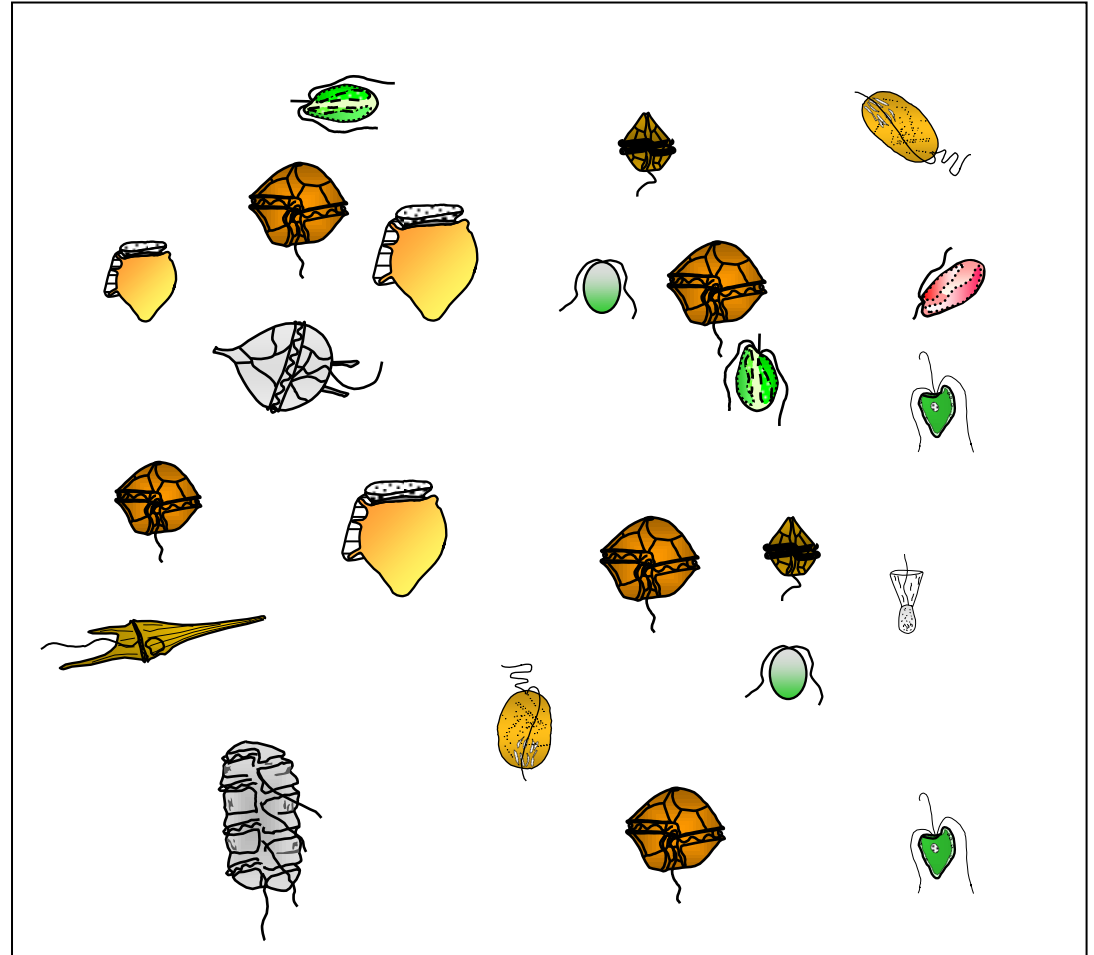
# HABs and phycotoxins: a general view and impacts on Chile

Bernd Krock

Alfred Wegener Institut-Helmholtz Zentrum für Polar- und Meeresforschung  
Am Handelshafen 12. 27576 Bremerhaven, Germany

Two aspects:

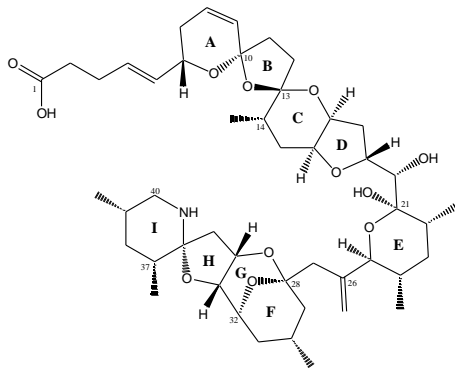
1. Shellfish poisoning syndromes
2. Ichthyotoxic (fishkilling) HAB Events



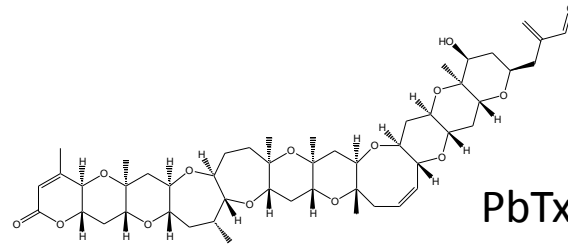
# Hab Events



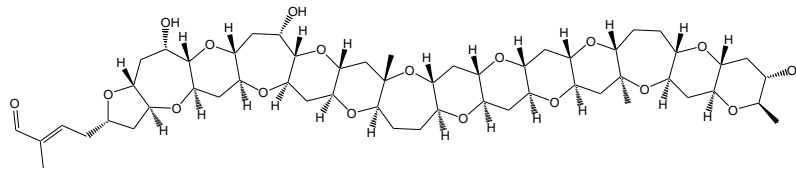
ALFRED-WEGENER-INSTITUT  
HELMHOLTZ-ZENTRUM FÜR POLAR-  
UND MEERESFORSCHUNG



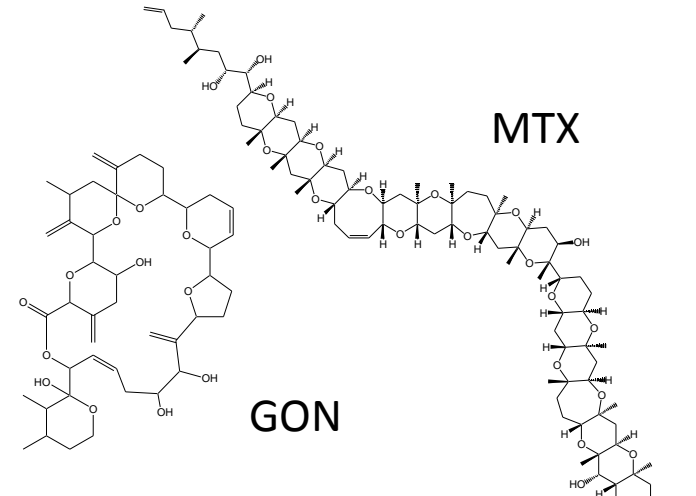
AZA



PbTx

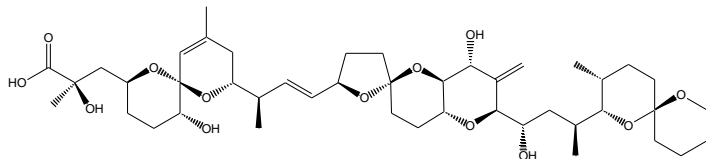


CTX

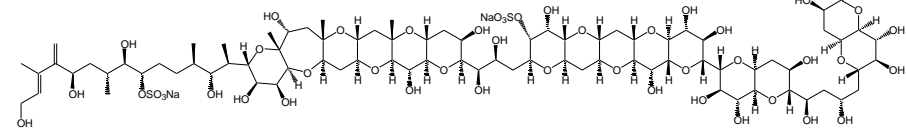


GON

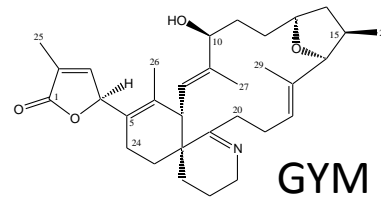
MTX



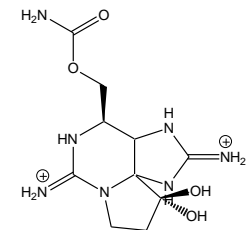
DTX



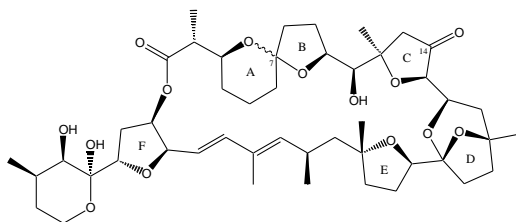
YTX



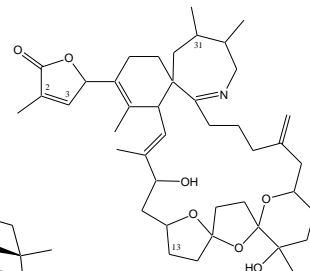
GYM



PST



PTX



SPX

Toxic effects on vertebrates (incl. humans):

Amnesic shellfish poisoning	ASP
-----------------------------	-----

Paralytic shellfish poisoning	PSP
-------------------------------	-----

Diarrhetic shellfish poisoning	DSP
--------------------------------	-----

Neurotoxic shellfish poisoning	NSP
--------------------------------	-----

Spiroimine shellfish poisoning	SSP
--------------------------------	-----

Ciguatera fish poisoning	CFP
--------------------------	-----

Azaspiracid shellfish poisoning	AZP
---------------------------------	-----

## Why select SSP and AZP?

- **Belong to the least researched syndromes**
- **Have an enormous toxin variability**
- **High potential of future risks**

## Spiroimines

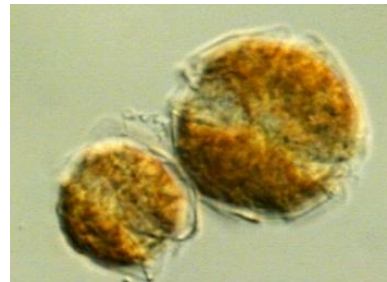
### Spirolide toxicity:

LD<sub>50</sub> (ip) in mice: 40 µg/kg

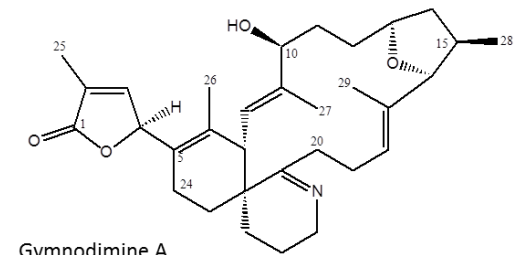
Oral toxicity: 1 mg/kg

### Gymnodimine toxicity:

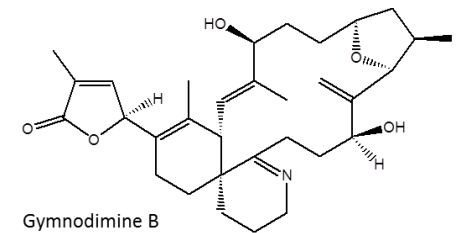
LD<sub>50</sub> (ip) in mice: 96 µg/kg



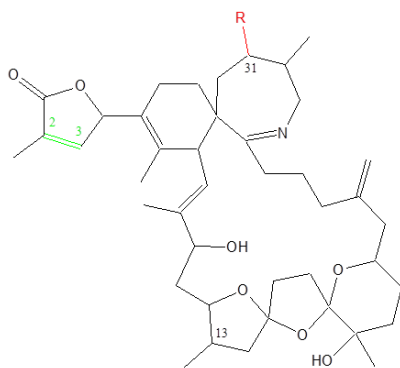
*Alexandrium ostenfeldii*



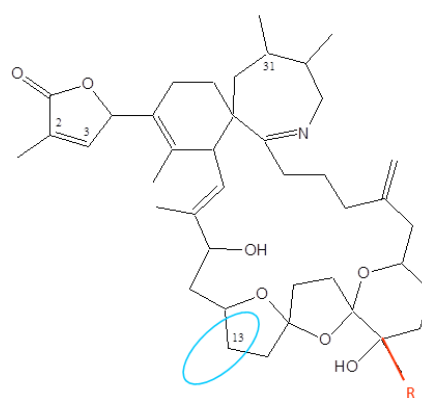
Gymnodimine A



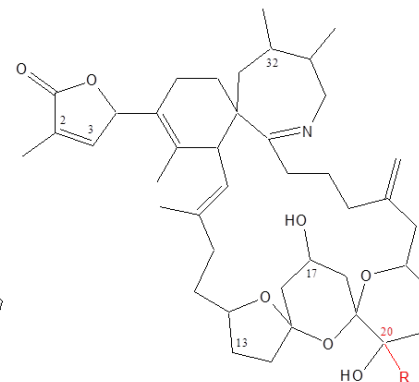
Gymnodimine B



Spirolide A: R = H, Δ<sup>2,3</sup>  
 B: R = H  
 C: R = Me, Δ<sup>2,3</sup>  
 D: R = Me



13-DesMe Spirolide C: R = Me  
 13,19-DidesMe Spirolide C: R = H



Spirolide G: R = H  
 20-Me Spirolide G: R = Me

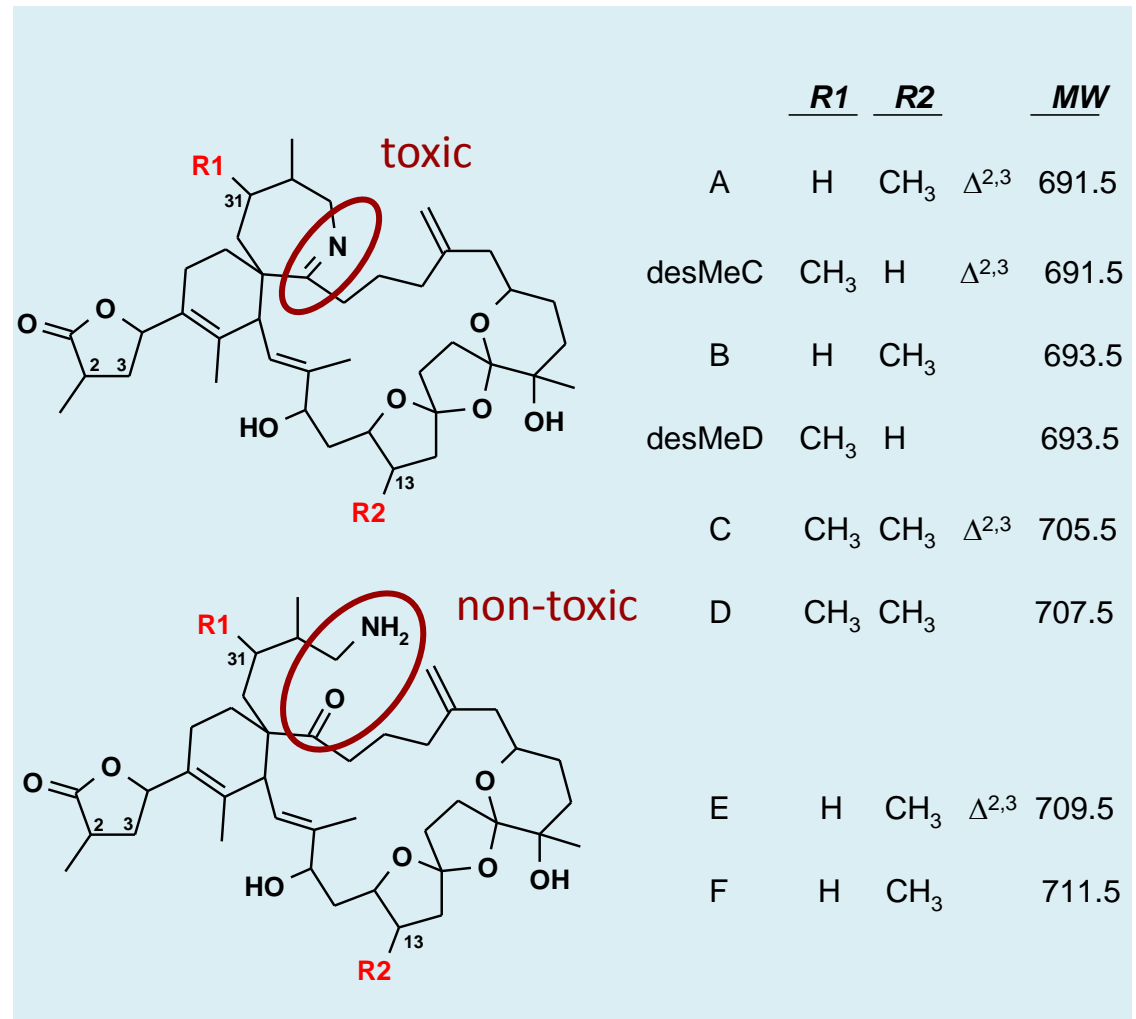
## Spirolides:

- macrocyclic imines
- structural similarity to pinnatoxins & gymnodimines
- pharmacologically active/inactive forms

## Mode of Action:

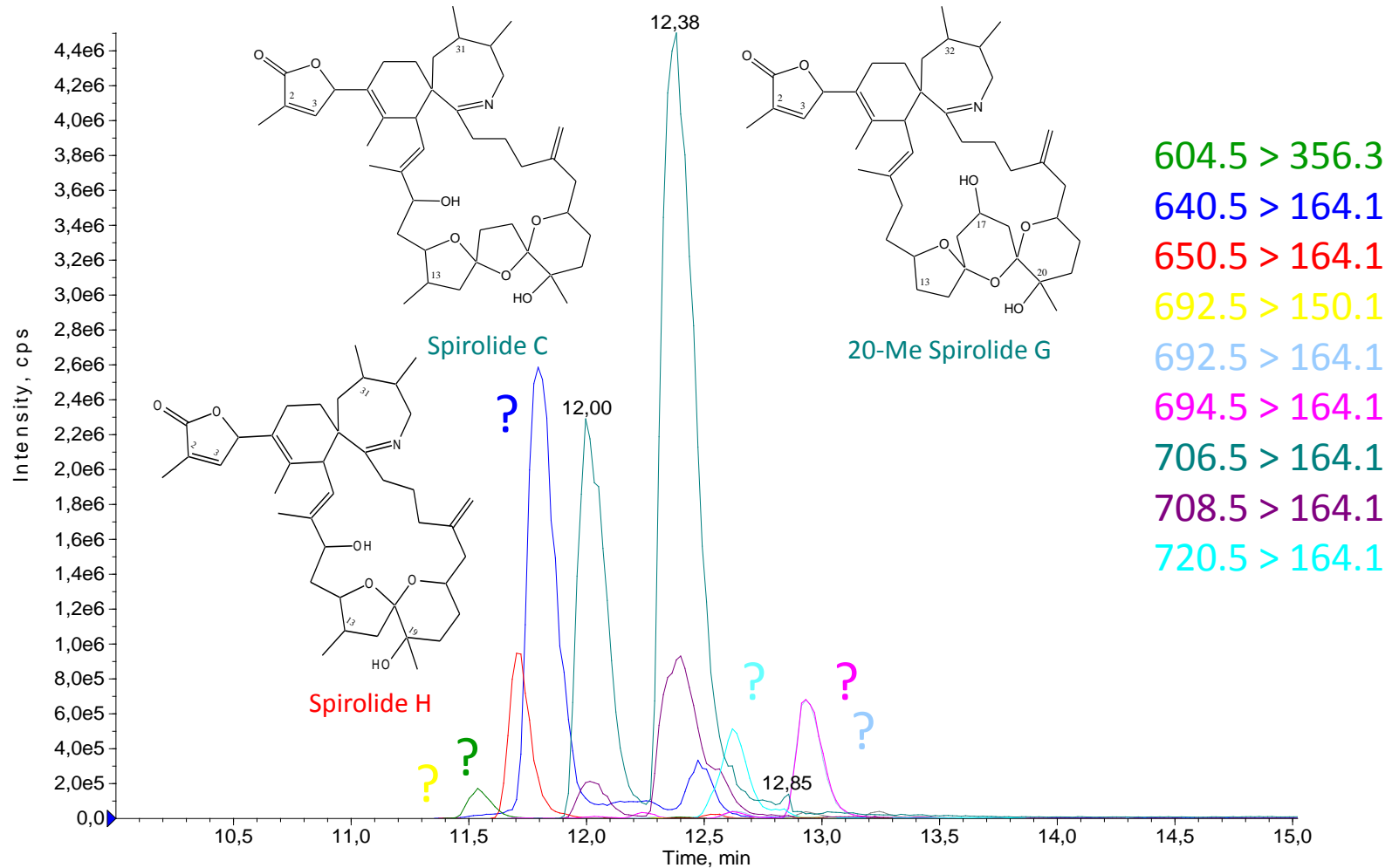
anticholinergic activity by blocking of muscarinic acetylcholin receptors

=> Paralysis of the parasympathetic nervous system (few toxicological studies)



# SSP high toxin variability within one strain

*Alexandrium ostenfeldii* (Ship Harbor, NS, Canada, AOSH2)



# SSP high toxin variability within populations



## Alexandrium ostenfeldii (Disko Island, Greenland)

	Stat	SPX-1	C	20-meG	H	Cp 1	Cp 2	Cp 3	Cp 4	Cp 5	Cp 6	Cp 7	Cp 8
P1 D5	506	-	77.1	16.8	-	-	-	-	-	-	3.4	2.7	-
P1 H10	516	0.7	-	84.3	-	0.1	27.5	0.3	-	0.8	-	-	13.7
P2 E3	516	31.2	-	-	41.3	-	-	-	-	-	-	-	-
P2 E4	516	19.2	-	-	-	1.2	-	-	-	70.1	-	-	9.4
P2 F2	516	5.1	63.6	-	6.7	0.1	4.1	-	-	10.0	-	3.8	-
P2 F3	516	-	82.9	17.1	-	-	-	-	-	-	-	-	-
P2 F4	516	2.7	57.3	39.7	-	-	-	-	-	-	0.1	-	-
P2 F7	516	1.4	31.1	-	25.1	1.2	10.0	-	-	29.0	-	-	2.3
P2 G2	516	0.2	40.2	18.4	7.1	-	11.1	-	-	11.0	-	-	11.3
P2 G9	516	-	100.0	-	-	-	-	-	-	-	-	-	-
P2 H4	516	-	95.4	4.6	-	-	-	-	-	-	-	-	-
P2 H8	516	2.4	-	89.0	-	-	-	-	-	0.3	-	-	8.2
P3 F1	516	0.2	-	81.7	-	-	0.1	0.2	-	1.0	-	-	16.9
P4 C6	516	-	50.3	49.7	-	-	-	-	-	-	-	-	-
P4 E3	516	1.2	96.3	2.2	-	-	-	0.3	-	-	-	-	-
P4 D8	516	-	31.4	-	14.1	-	-	9.0	36.1	9.4	-	-	-
P4 F10	516	-	68.6	20.0	0.2	-	-	-	-	-	-	-	11.3
P4 G2	516	-	-	100.0	-	-	-	-	-	-	-	-	-
P3 A12	516	-	52.2	-	47.8	-	-	-	-	-	-	-	-
P2 H2	516	-	-	92.5	-	-	-	0.9	-	-	-	-	6.6
P2 G3	516	18.1	1.3	0.1	-	-	-	-	-	72.7	-	-	7.8
P3 E4	516	0.2	99.6	-	-	-	-	0.2	-	-	-	-	-
P4 F4	516	-	77.4	19.6	0.3	-	-	0.4	-	2.1	-	-	-
P1 F5	524	0.1	79.5	19.8	-	-	-	0.2	-	-	0.1	0.2	-
P1 F7	524	0.1	78.3	20.9	-	-	0.1	0.4	-	-	0.1	-	-
P1 F8	524	-	92.1	6.6	-	-	0.1	0.4	-	-	0.2	0.5	-
P1 F9	524	0.2	64.7	33.2	-	-	0.1	1.4	-	-	0.3	-	-
P1 F10	524	0.1	68.4	30.5	-	-	0.1	0.9	-	-	0.1	-	-
P1 F11	524	-	87.3	6.4	-	-	-	-	-	-	-	5.4	-
P1 G3	524	0.6	92.3	6.6	-	-	-	0.6	-	-	-	-	-
P1 G5	524	0.7	76.1	21.7	0.4	-	-	0.4	-	-	0.3	0.4	-
P1 G11	524	0.2	84.2	14.2	0.5	-	-	0.3	-	-	0.3	0.3	-
P1 G8	524	-	88.6	6.5	-	-	-	-	-	-	-	4.9	-
P1 F6	524	0.1	85.1	13.8	-	-	0.1	0.1	-	-	0.1	0.3	-
P1 F4	524	0.1	77.2	22.3	-	-	-	-	-	-	0.1	0.2	-
P1 G6	524	-	70.5	28.0	0.3	-	-	0.6	-	-	0.2	0.3	-

Tillmann et al. (2014)  
*Harmful Algae* 39, 259-270.



# SSP high toxin variability within populations

## known variants

Mass transition	toxin
508>490	GYM A
522>504	12-me GYM A
650>164	H
652>164	I
678>164	13,19-didesme C
692>164	13-desme C, G, undescribed
692>178	27-oxo-13,19-didesme C
692>150	A, undescribed
694>164	13-desme D, undescribed, pinnatoxin G
694>180	27-hydroxy-13,19-didesme C
694>150	B
706>164	C, 20-me G
706>164	27-hydroxy-13-desme C
708>164	D
766>164	pinnatoxin F
784>164	pinnatoxin E

## undescribed variants

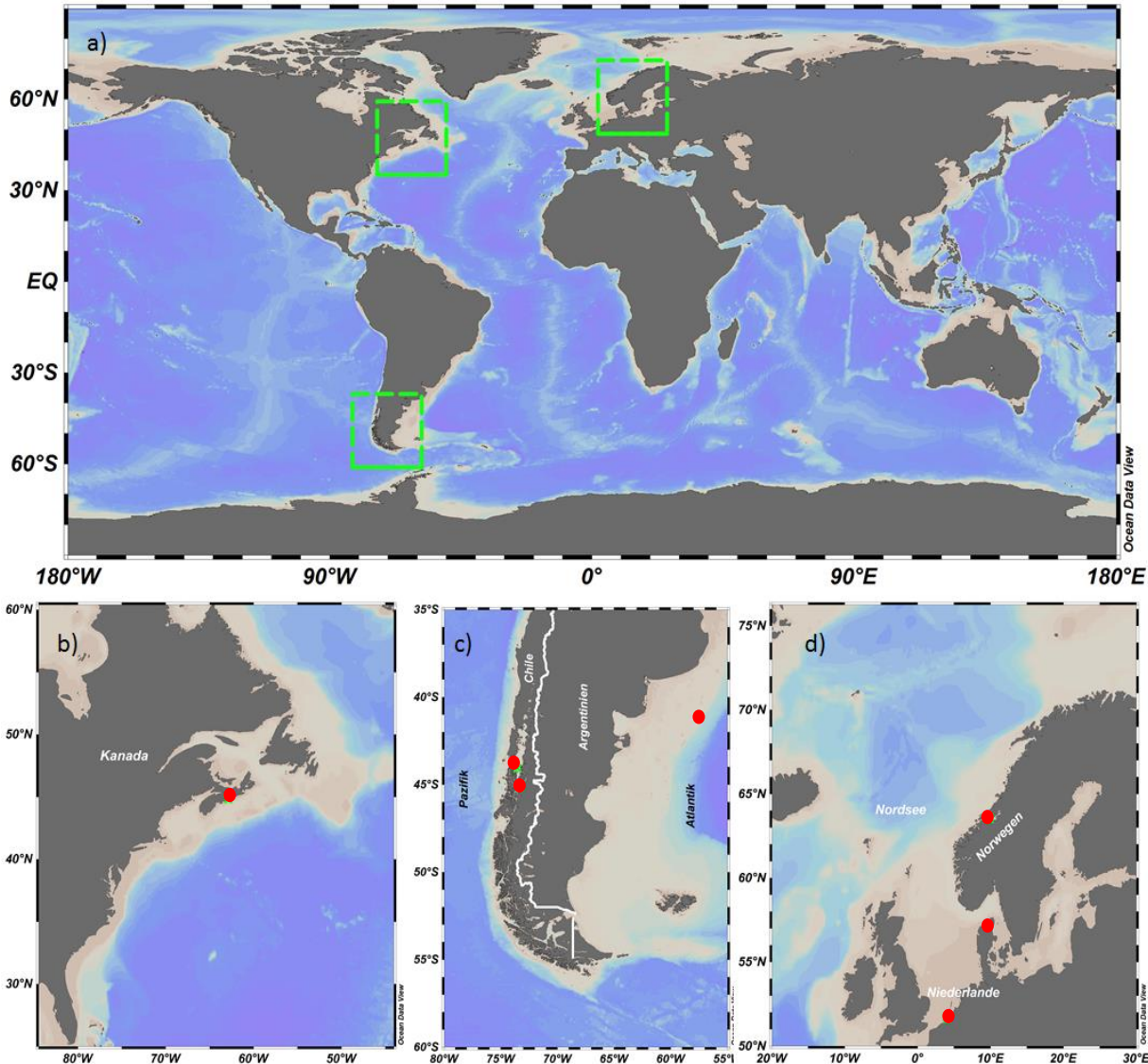
Mass transition	spirolide
640>164	undescribed
644>164	undescribed
658>164	undescribed
658>150	undescribed
674>164	undescribed
678>150	undescribed
692>150	A, undescribed
696>164	undescribed
698>164	undescribed
710>164	undescribed
710>150	undescribed
720>164	undescribed
722>164	undescribed
722>180	undescribed

Tillmann et al. (2014) Harmful Algae 39, 259-270.

# SSP high toxin variability among populations



ALFRED-WEGENER-INSTITUT  
HELMHOLTZ-ZENTRUM FÜR POLAR-  
UND MEERESFORSCHUNG

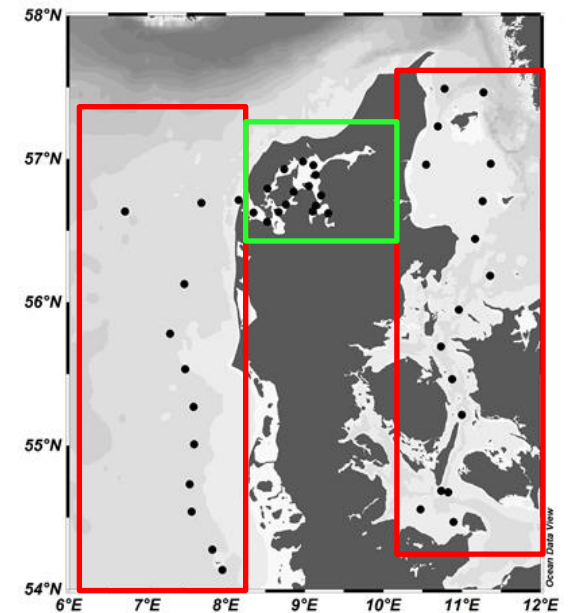


Krohn 2016 (2012) Bachelor thesis,  
University of Lübeck, Germany

# SSP high toxin variability among populations

Strain	Salinity	Origin	PSP	SPX	GYM
AOSH2	> 30	Atlantic Canada	/	x	/
NX 56-07	21,1 - 33,3	Norway	/	x	x
NX 56-10	21,1 - 33,4	Norway	/	x	x
NX 56-12	21,1 - 33,5	Norway	/	x	x
ND	33,3	North Sea*	/	x	/
ND	26,1	Limfjord DK*	/	x	x
ND	24,5	Kattegat*	/	x	/
OKNL 42	8 - 21	The Netherlands	x	x	x
AOICW	32,5	Chile	x	x	/
AOA32-2	28,2	Chile	x	x	/
H1G8	33,6	Argentina	x	x	/
H3D4	33,6	Argentina	x	x	/
H2-A4	33,6	Argentina	x	x	/

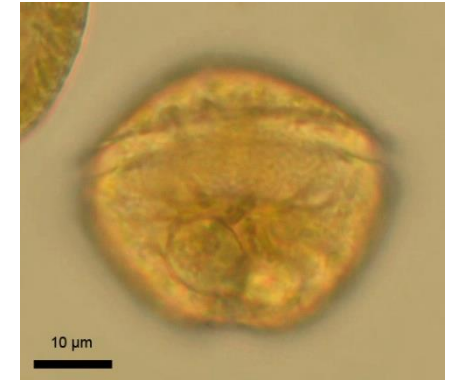
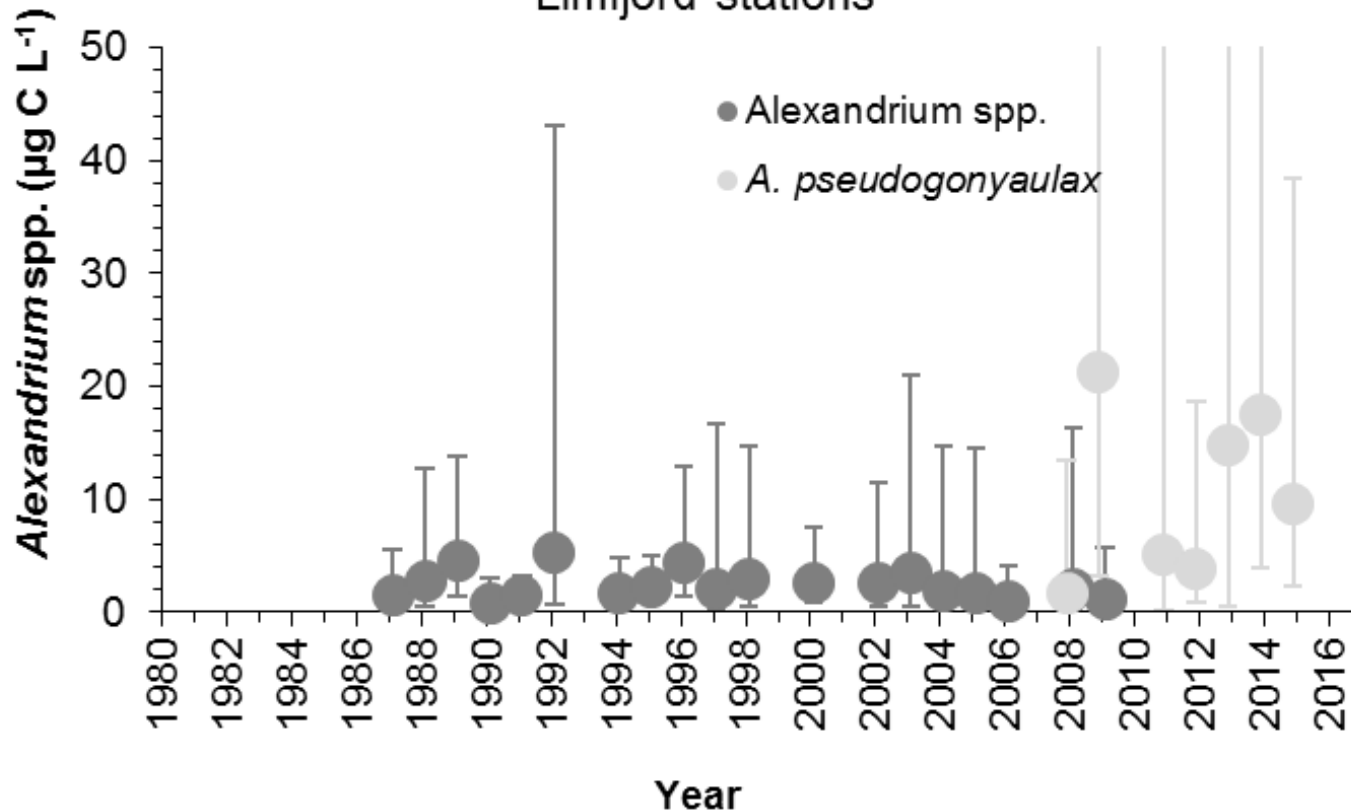
\* Field samples





# Dinoflagellate community shift

Summer biomass of *Alexandrium* spp. at the NOVANA  
Limfjord stations



*Alexandrium pseudogonyaulax*

Producer of Goniodomins

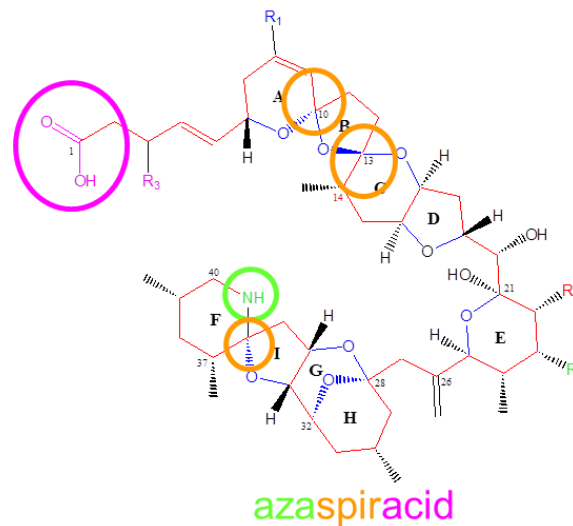
1995: 8 people in the Netherlands became ill after consumption of Irish mussels (*Mytilus edulis*) harvested at Killary Harbour (Ireland). Symptoms were like DSP intoxication, but DSP toxins were hardly present in the mussels (MacMahon & Silke, 1996: Harmful Algae News, 14, 2)

## Toxicity:

LD<sub>50</sub> AZA-1: 0.2 µg/kg (mice)

## Mode of Action:

- effects of AZA-1 on the arrangement of F-actin  
 ⇒ concurrent loss of pseudopodia, cytoplasmic extensions that function in mobility and chemotaxis; effects on cytoskeleton
- Increases cytosolic calcium levels in lymphocytes
- diarrheagenic, tumorigenic



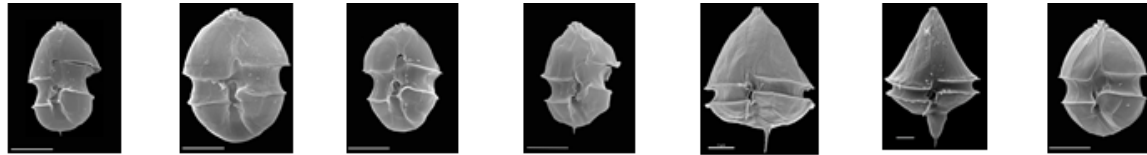
Polyketide:

linear carbon skeleton  
 with cyclic ether  
 bridges

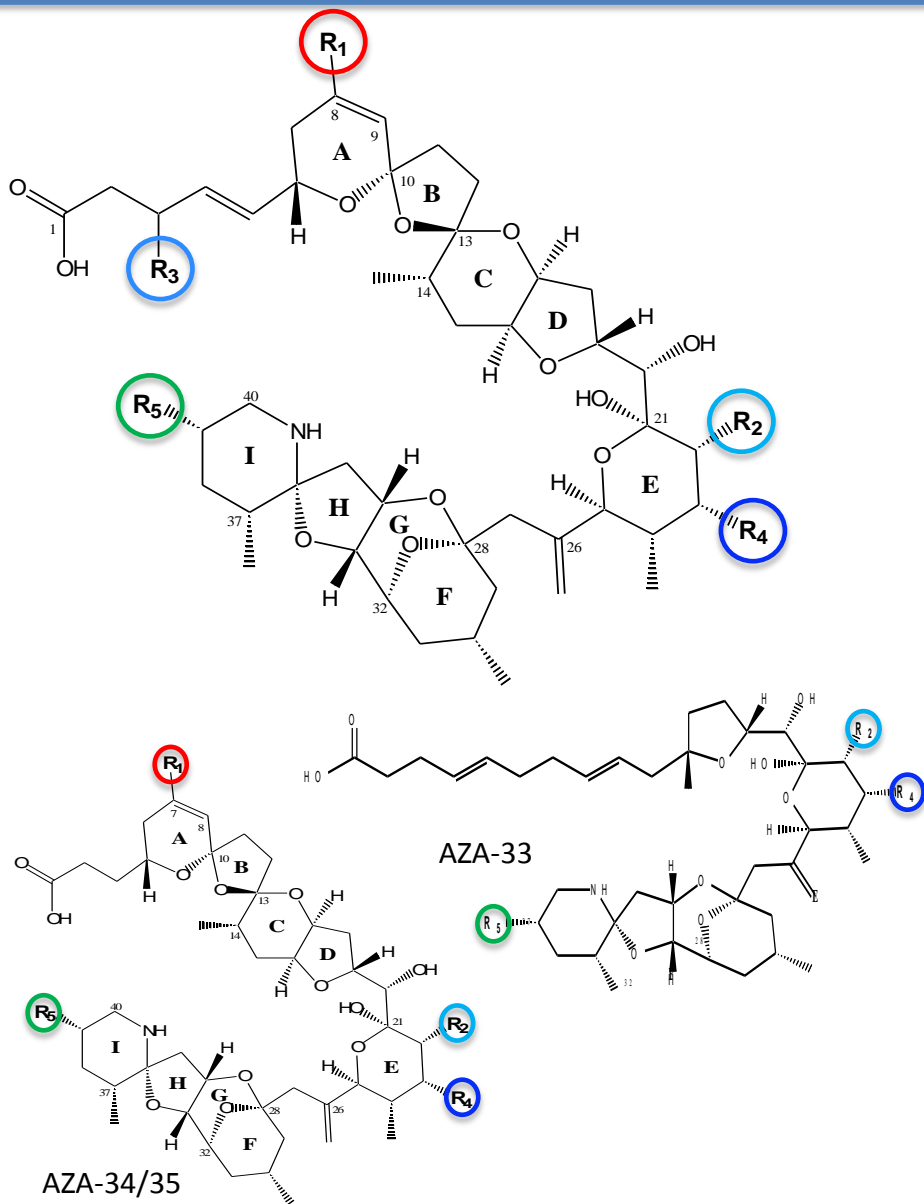
amino function  
 chemical nomenclature:  
 aza = secondary amine

spiro function

acid



	<i>Azadinium spinosum</i>	<i>Azadinium obesum</i>	<i>Azadinium poporum</i>	<i>Azadinium polongum</i>	<i>A. caudatum</i> var. <i>margalefii</i>	<i>A. caudatum</i> var. <i>caudatum</i>	<i>Amphidoma languida</i>
size	13.8 x 8.8	15.3 x 11.7	13.0 x 9.8	13.0 x 9.7	31.1 x 22.4	41.7 x 28.7	13.9. x 11.9
length/width ratio	1.6	1.3	1.3	1.3	1.2	1.2	1.2
Pyrenoid	1; epicone	-	Several (4?), epi- and hypocone	-	-	-	1; epicone
Spine	+	-	-	+	<b>Short horn, long spine</b>	<b>Long horn, short spine</b>	
Ventral pore	+	+	-	+	-	+	+
Pore on po-plate	-	-	+	-	+	-	-
Shape pore-plate	<b>round- elipsoid</b>	<b>round- elipsoid</b>	<b>round- elipsoid</b>	<b>elongated</b>	<b>round- elipsoid</b>	<b>round- elipsoid</b>	<b>round- elipsoid</b>
Plate 1'' in contact to plate 1a	+	-	+	+	+	+	+
Plate 6'' in contact to plate 3a	-	-	-	-	+	+	-
<b>Azaspiracids</b>	<b>+</b>	<b>-</b>	<b>+</b>	<b>-</b>	<b>-</b>	<b>-</b>	<b>+</b>



Toxin	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>	R <sub>4</sub>	R <sub>5</sub>	Δ <sub>7,8</sub>	[M+H] <sup>+</sup>
AZA-1	H	CH <sub>3</sub>	H	H	CH <sub>3</sub>	✓	842
AZA-2	CH <sub>3</sub>	CH <sub>3</sub>	H	H	CH <sub>3</sub>	✓	856
AZA-3	H	H	H	H	CH <sub>3</sub>	✓	828
AZA-4	H	H	OH	H	CH <sub>3</sub>	✓	844
AZA-5	H	H	H	OH	CH <sub>3</sub>	✓	844
AZA-6	CH <sub>3</sub>	H	H	H	CH <sub>3</sub>	✓	842
AZA-7	H	CH <sub>3</sub>	OH	H	CH <sub>3</sub>	✓	858
AZA-8	H	CH <sub>3</sub>	H	OH	CH <sub>3</sub>	✓	858
AZA-9	CH <sub>3</sub>	H	OH	H	CH <sub>3</sub>	✓	858
AZA-10	CH <sub>3</sub>	H	H	OH	CH <sub>3</sub>	✓	858
AZA-11	CH <sub>3</sub>	CH <sub>3</sub>	OH	H	CH <sub>3</sub>	✓	872
AZA-33	-	CH <sub>3</sub>	H	H	CH <sub>3</sub>	-	716
AZA-34	H	CH <sub>3</sub>	-	H	CH <sub>3</sub>	✓	816
AZA-35	CH <sub>3</sub>	CH <sub>3</sub>	-	H	CH <sub>3</sub>	✓	830
AZA-36	CH <sub>3</sub>	CH <sub>3</sub>	OH	H	H	✓	858
AZA-37	H	CH <sub>3</sub>	OH	H	H	-	846
AZA-38	nd	nd	nd	nd	H	nd	830
AZA-39	nd	nd	nd	nd	H	nd	816
AZA-40	nd	nd	nd	nd	H	nd	842
AZA-41	nd	nd	nd	nd	CH <sub>3</sub>	nd	854
AZA-42	nd	nd	nd	nd	CH <sub>3</sub>	nd	870



## Planktonic strain AZAs

AZA-1	AZA-41
AZA-2	AZA-42
epi-AZA-7	AZA-43
AZA-11	AZA-50
AZA-33	AZA-51
AZA-34	AZA-52
AZA-35	AZA-53
AZA-36	AZA-54
AZA-37	AZA-55
AZA-38	AZA-56
AZA-39	AZA-57
AZA-40	AZA-58

## AZA shellfish metabolites of AZA-1 and -2

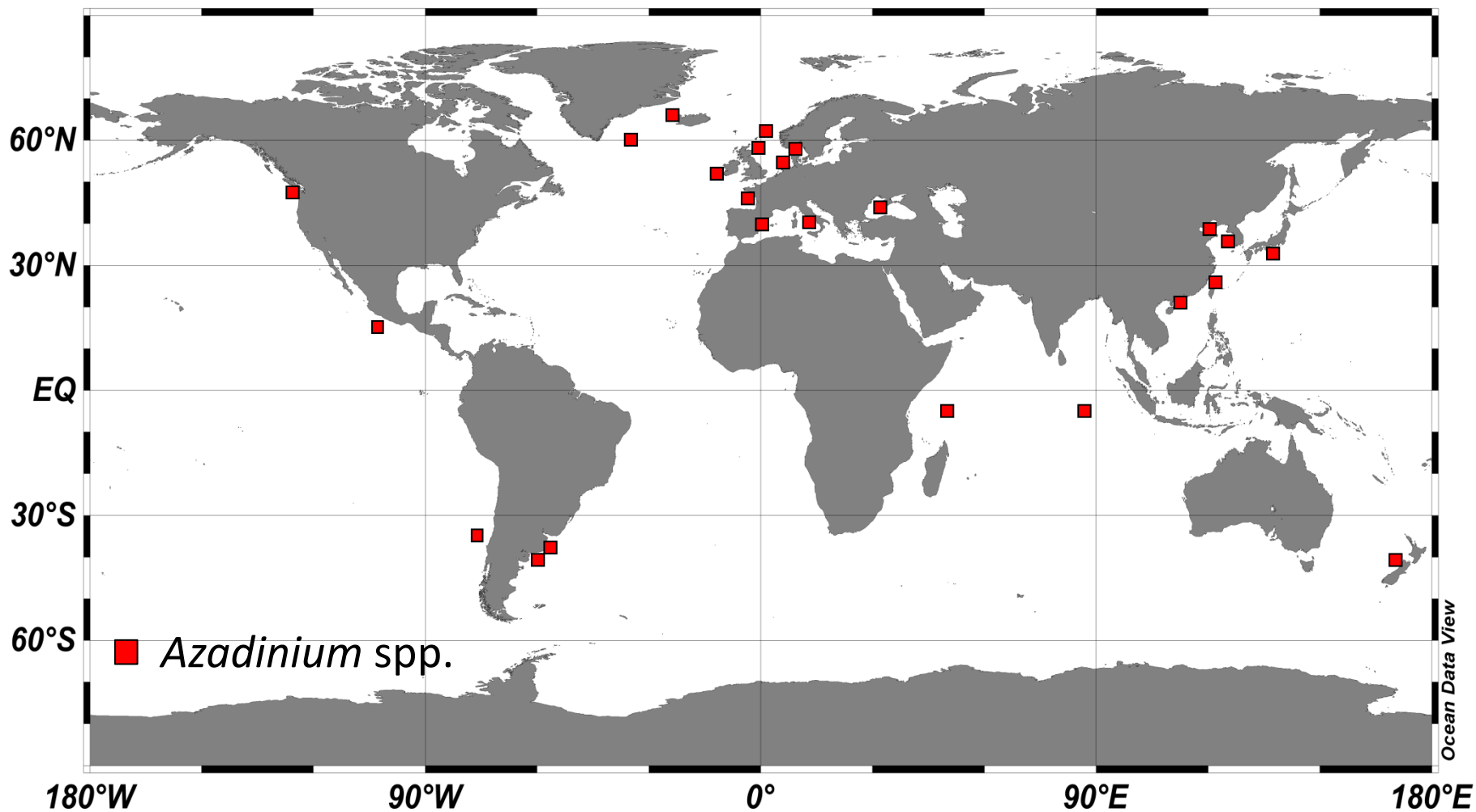
AZA-3	AZA-14	AZA-25	AZA-47
AZA-4	AZA-15	AZA-26	AZA-48
AZA-5	AZA-16	AZA-27	AZA-49
AZA-6	AZA-17	AZA-28	
AZA-7	AZA-18	AZA-29	
AZA-8	AZA-19	AZA-30	
AZA-9	AZA-20	AZA-31	
AZA-10	AZA-21	AZA-32	
AZA-11	AZA-22	AZA-44	
AZA-12	AZA-23	AZA-45	
AZA-13	AZA-24	AZA-46	

Two AZAs of phytoplankton origin result in 36 shellfish metabolites!

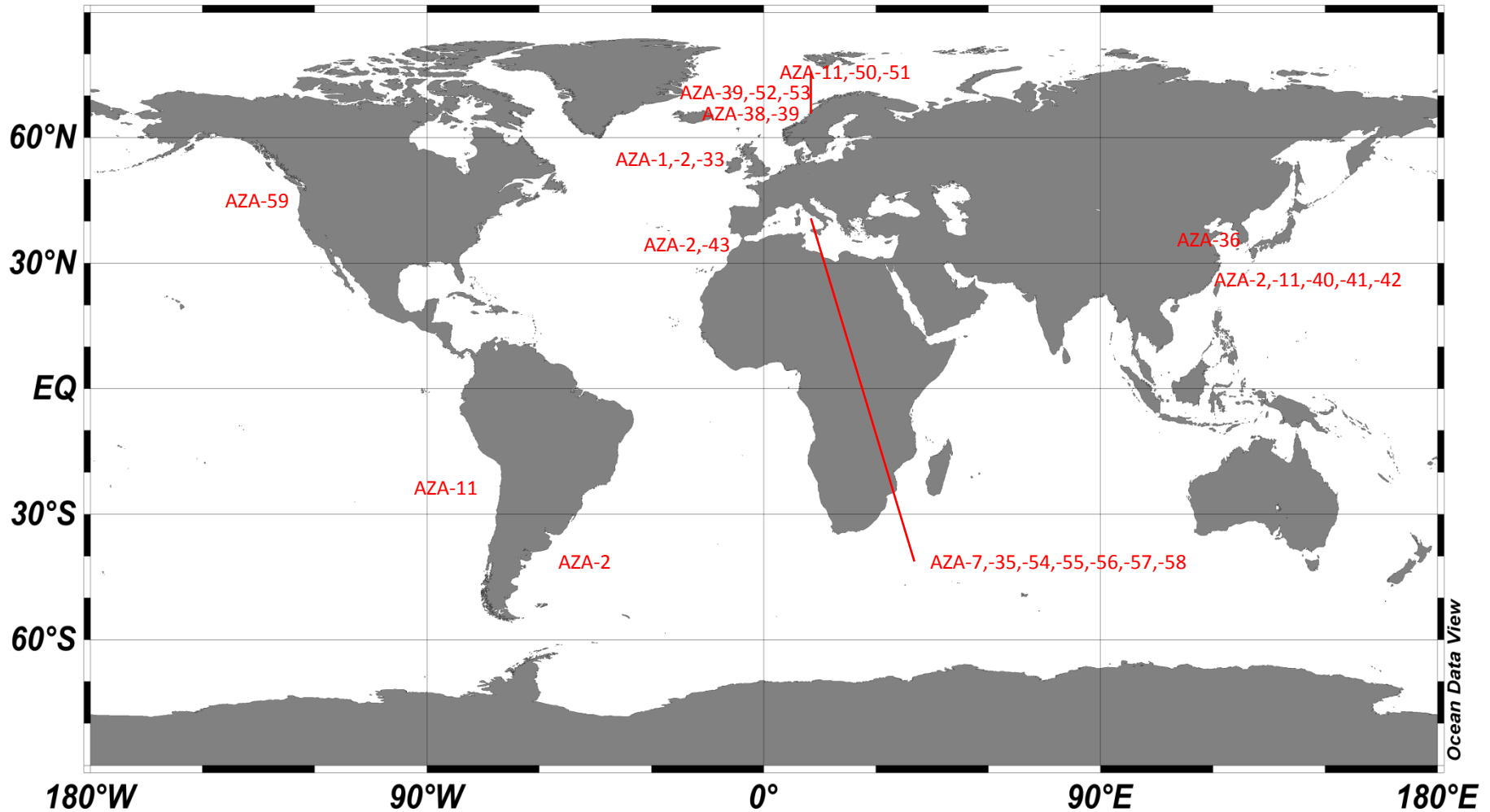
AZA	R1	R2	R3	R4	R5	R6	R7	[M+H] <sup>+</sup>	Frag.Type	origin	status	reference
AZA1	H	H	H	CH3	H	CH3	CH3	842,5	362 - 262	A. spinosum	phycotoxin	Rehmann et al. 2008
AZA2	H	CH3	H	CH3	H	CH3	CH3	856,5	362 - 262	spin/pop/lang	phycotoxin	Rehmann et al. 2008
AZA3	H	H	H	H	H	CH3	CH3	828,5	362 - 262	shellfish	metabolite	Rehmann et al. 2008
AZA4	OH	H	H	H	H	CH3	CH3	844,5	362 - 262	shellfish	metabolite	Rehmann et al. 2008
AZA5	H	H	H	H	OH	CH3	CH3	844,5	362 - 262	shellfish	metabolite	Rehmann et al. 2008
AZA6	H	CH3	H	H	H	CH3	CH3	842,5	362 - 262	shellfish	metabolite	Rehmann et al. 2008
AZA7	OH	H	H	CH3	H	CH3	CH3	858,5	362 - 262	shellfish	metabolite	Rehmann et al. 2008
epi-AZA7	OH	H	H	CH3	H	CH3	CH3	858,5	362 - 262	A. dexteroporum	phycotoxin	Rossi et al. 2017
AZA8	H	H	H	CH3	OH	CH3	CH3	858,5	362 - 262	shellfish	metabolite	Rehmann et al. 2008
AZA9	OH	CH3	H	H	H	CH3	CH3	858,5	362 - 262	shellfish	metabolite	Rehmann et al. 2008
AZA10	H	CH3	H	H	OH	CH3	CH3	858,5	362 - 262	shellfish	metabolite	Rehmann et al. 2008
AZA11	OH	CH3	H	CH3	H	CH3	CH3	872,5	362 - 262	Pop/shelf	phycotox, metaboli	Rehmann et al. 2008
AZA12	H	CH3	H	CH3	OH	CH3	CH3	872,5	362 - 262	shellfish	metabolite	Rehmann et al. 2008
AZA13	OH	H	H	H	OH	CH3	CH3	860,5	362 - 262	shellfish	metabolite	Rehmann et al. 2008
AZA14	OH	H	H	CH3	OH	CH3	CH3	874,5	362 - 262	shellfish	metabolite	Rehmann et al. 2008
AZA15	OH	CH3	H	H	OH	CH3	CH3	874,5	362 - 262	shellfish	metabolite	Rehmann et al. 2008
AZA16	OH	CH3	H	CH3	OH	CH3	CH3	888,5	362 - 262	shellfish	metabolite	Rehmann et al. 2008
AZA17	H	H	H	COOH	H	CH3	CH3	872,5	362 - 262	shellfish	metabolite	Rehmann et al. 2008
AZA18												Rehmann et al. 2008
AZA19	H	CH3	H	COOH	H	CH3	CH3	886,5	362 - 262	shellfish	metabolite	Rehmann et al. 2008
AZA20												Rehmann et al. 2008
AZA21	OH	H	H	COOH	H	CH3	CH3	888,5	362 - 262	shellfish	metabolite	Rehmann et al. 2008
AZA22												Rehmann et al. 2008
AZA23	OH	CH3	H	COOH	H	CH3	CH3	902,5	362 - 262	shellfish	metabolite	Rehmann et al. 2008
AZA24												Rehmann et al. 2008
AZA25	H	H	H	H	H	CH3	CH3	810,5	362 - 262	shellfish	metabolite	Kilkoyne et al. unpubl.
AZA26	H	H	H	H	O	CH3	CH3	824,5	362 - 262	shellfish	metabolite	Kilkoyne et al. unpubl.
AZA27	H	CH3	H	H	H	CH3	CH3	824,5	362 - 262	shellfish	metabolite	Kilkoyne et al. unpubl.
AZA28	H	CH3	H	H	O	CH3	CH3	838,5	362 - 262	shellfish	metabolite	Kilkoyne et al. unpubl.
AZA29	H	H	CH3	H	H	CH3	CH3	842,5	362 - 262	shellfish	artefact	Rehmann et al. 2008
AZA30	H	H	CH3	CH3	H	CH3	CH3	856,5	362 - 262	A. spinosum	artefact	Rehmann et al. 2008
AZA31												Rehmann et al. 2008
AZA32	H	CH3	CH3	CH3	H	CH3	CH3	870,5	362 - 262	A. spinosum	artefact	Rehmann et al. 2008
AZA33	-	-	H	CH3	H	CH3	CH3	716,5	362 - 262	A. spinosum	phycotoxin	Kilkoyne et al. 2014
AZA34	-	H	H	CH3	H	CH3	CH3	816,5	362 - 262	A. spinosum	phycotoxin	Kilkoyne et al. 2014
AZA35	-	CH3	H	CH3	H	CH3	CH3	830,5	362 - 262	A. spin/A. dextero	phycotoxin	Kilkoyne et al. 2014, Rossi et al. 2017
AZA36	OH	CH3	H	CH3	H	H	CH3	858,5	348-248	A. poporum	phycotoxin	Krock et al. 2015
AZA37	OH	H	H	CH3	H	H	CH3	846,5	348-248	A. poporum	phycotoxin	Krock et al. 2015
AZA38	nd	nd	nd	nd	nd	nd	nd	830,5	348-248	A. languida	phycotoxin	Krock et al. 2012
AZA39	nd	nd	nd	nd	nd	nd	nd	816,5	348-248	A. languida	phycotoxin	Krock et al. 2012
AZA40	H	CH3	H	CH3	H	H	CH3	842,5	348-248	A. poporum	phycotoxin	Krock et al. 2014
AZA41	H	CH3	H	CH3	H	CH3	CH3	854,5	360-260	A. poporum	phycotoxin	Krock et al. 2014
AZA42	OH	CH3	H	CH3	H	CH3	CH3	856,5	360-260	A. poporum	phycotoxin	Krock & Tillmann, unpubl.
AZA43	-	H	H	CH3	H	H	CH3	828,5	360-260	A. languida	phycotoxin	Tillmann et al. 2017
AZA44	H	H	H	COOH	OH	CH3	CH3	888,5	362-262	shellfish	metabolite	Kilkoyne et al. 2015
AZA45	H	CH3	H	COOH	OH	CH3	CH3	902,5	362-262	shellfish	metabolite	Kilkoyne et al. 2015
AZA46	OH	H	H	COOH	OH	CH3	CH3	904,5	362-262	shellfish	metabolite	Kilkoyne et al. 2015
AZA47	OH	CH3	H	COOH	OH	CH3	CH3	918,5	362-262	shellfish	metabolite	Kilkoyne et al. 2015
AZA48	OH	H	H	H	H	CH3	CH3	826,5	362-262	shellfish	metabolite	Kilkoyne unpublished
AZA49	OH	CH3	H	H	H	CH3	CH3	840,5	362-262	shellfish	metabolite	Kilkoyne unpublished
AZA50	H	CH3	H	CH3	H	CH3	H	842,5	348-262	A. spinosum	Phycotoxin	Krock & Tillmann, unpubl.
AZA51	OH	CH3	H	CH3	H	CH3	H	858,5	348-262	A. spinosum	Phycotoxin	Krock & Tillmann, unpubl.
AZA52	nd	nd	nd	nd	nd	nd	nd	830,5	348-248	A. languida	Phycotoxin	Krock & Tillmann, unpubl.
AZA53	nd	nd	nd	nd	nd	nd	nd	830,5	348-248	A. languida	Phycotoxin	Krock & Tillmann, unpubl.
AZA54	nd	nd	nd	nd	nd	nd	nd	870,5	362 - 262	A. dexteroporum	Phycotoxin	Rossi et al. 2017
AZA55	nd	nd	nd	nd	nd	nd	nd	868,5	362 - 262	A. dexteroporum	Phycotoxin	Rossi et al. 2017
AZA56	nd	nd	nd	nd	nd	nd	nd	884,5	362 - 262	A. dexteroporum	Phycotoxin	Rossi et al. 2017
AZA57	nd	nd	nd	nd	nd	nd	nd	826,5	362 - 262	A. dexteroporum	Phycotoxin	Rossi et al. 2017
AZA58	nd	nd	nd	nd	nd	nd	nd	828,5	362 - 262	A. dexteroporum	Phycotoxin	Rossi et al. 2017
AZA59	OH	H	H	CH3	H	CH3	CH3	860,5	362 - 262	A. poporum	Phycotoxin	Kim et al. 2017

Currently  
60 known AZAs

## Occurrence of *Azadinum* spp. & *Amphidoma languida*



## AZA profiles of species and strains of different origins



## ➤ AZPs en Chile

### *Azadinium spinosum*

López Rivera et al. 2010  
(ostiones, 2005-2006  
bahía Inglesa, Salada, Tongoy)  
ND-96 µg/kg (g.d.)

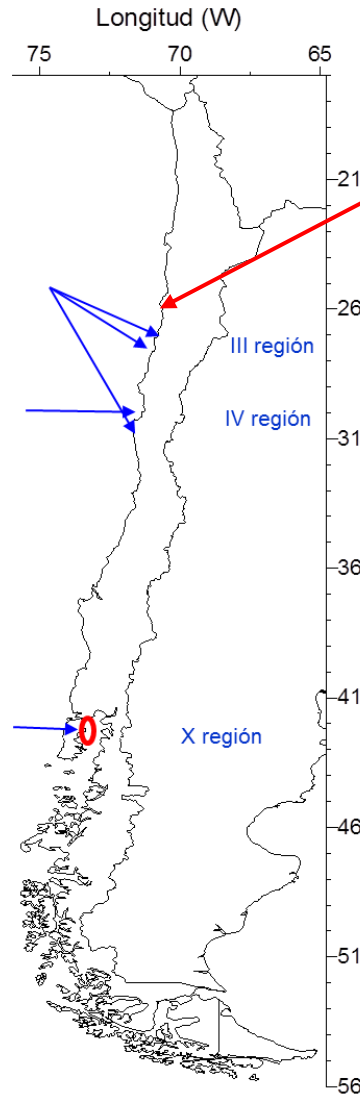
Álvarez et al. 2010  
(almejas, machas, 2008,  
bahía Coquimbo)  
LD=2.42 µg/kg

López Rivera et al. 2010  
choritos, 2005-2006,  
Mar int. Chiloé)  
18-31 µg/kg (c.e.)

Reports of AZA-1,  
-2 -3 and -6 in  
mollusks

Pizarro, G. pers. comm.

Límite normativo máximo  
(consumo humano)  
160 µg/kg

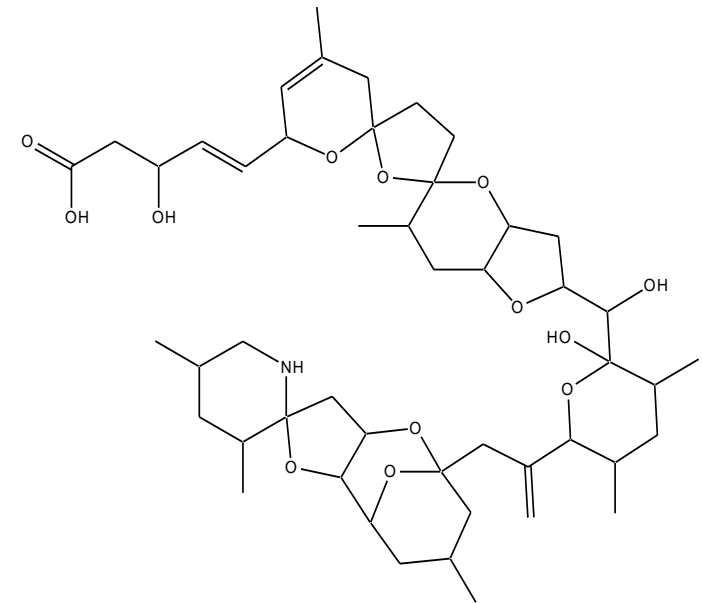


### *Azadinium poporum*

#### Chañaral area

Tillmann et al. (2017) J. Plankt. Res., 39, 350-367.

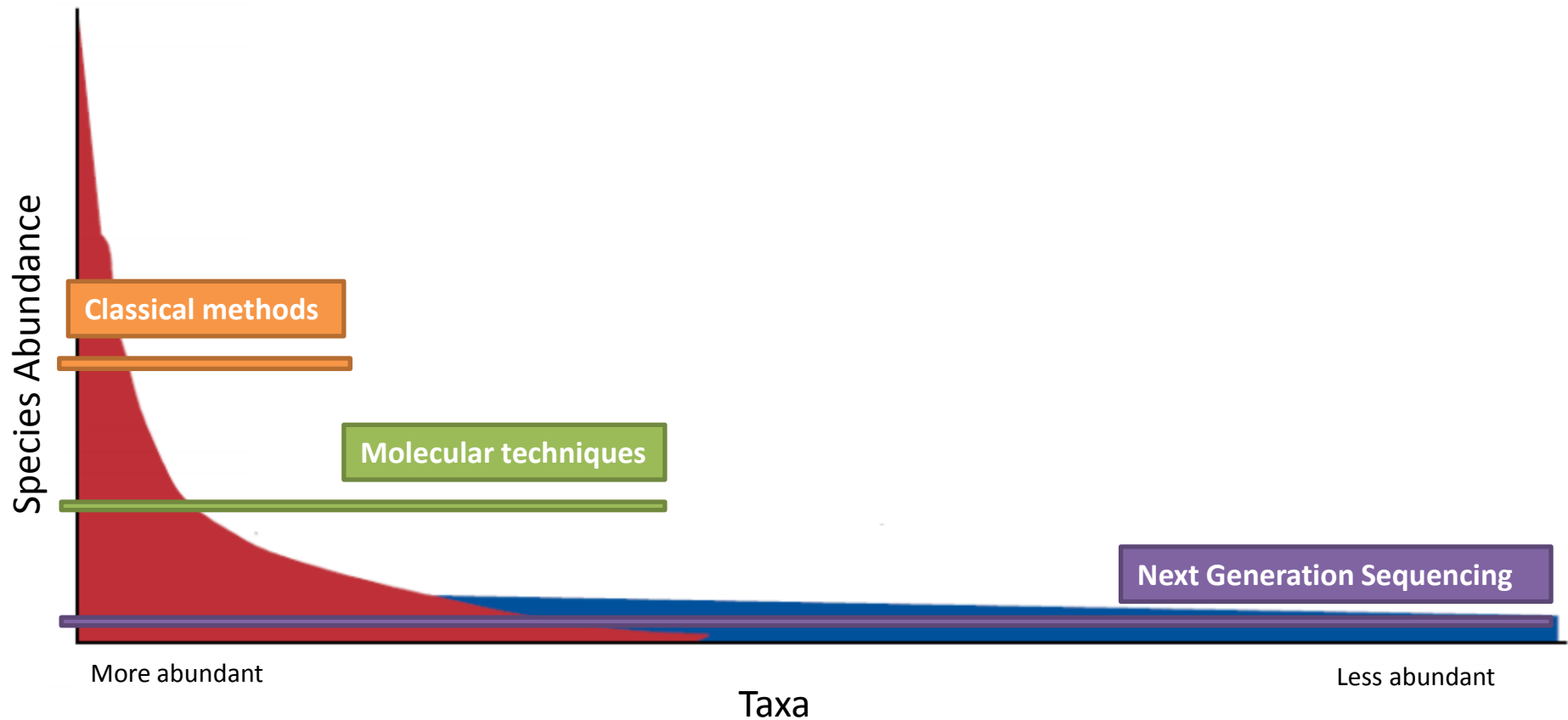
### AZA-11 (3-4 fg/cell) and AZA-11 phosphate



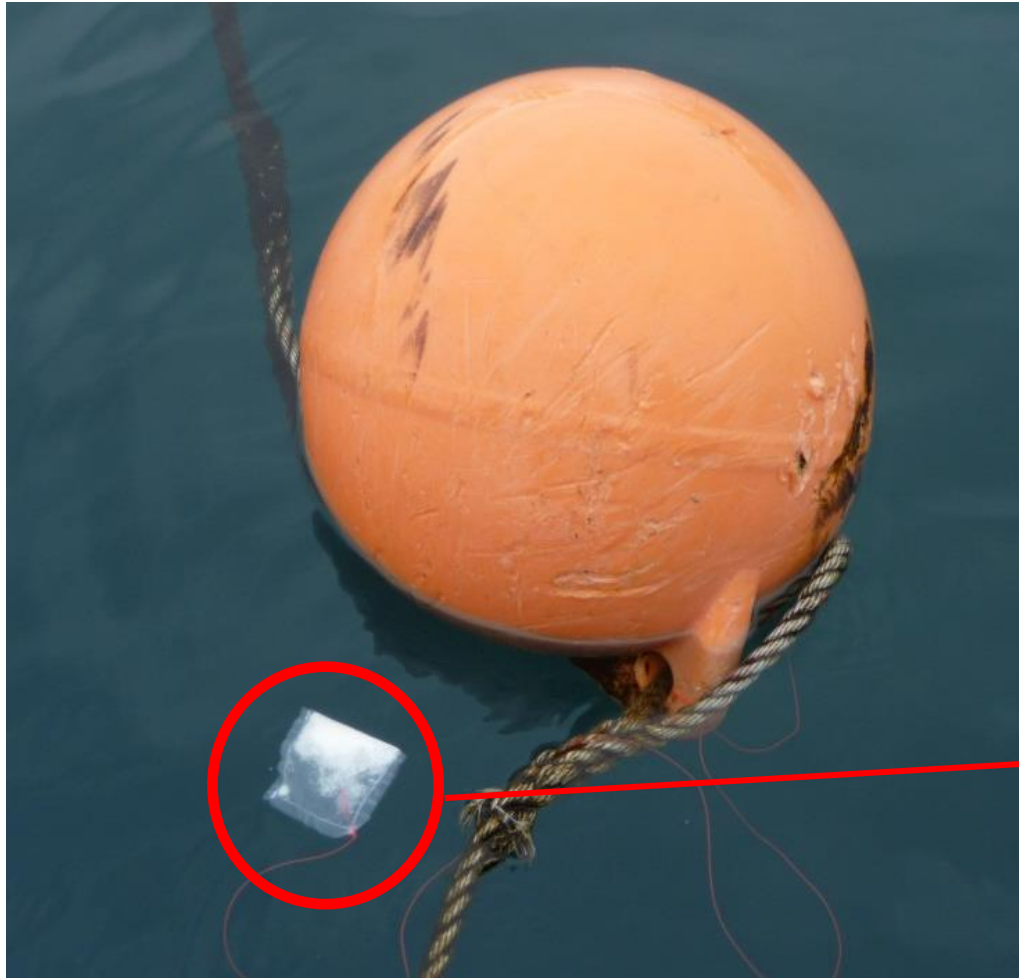
# Cryptic species

## Metagenomics of plankton

Moreno-Pino et al., submitted



# HAB species range expansion?

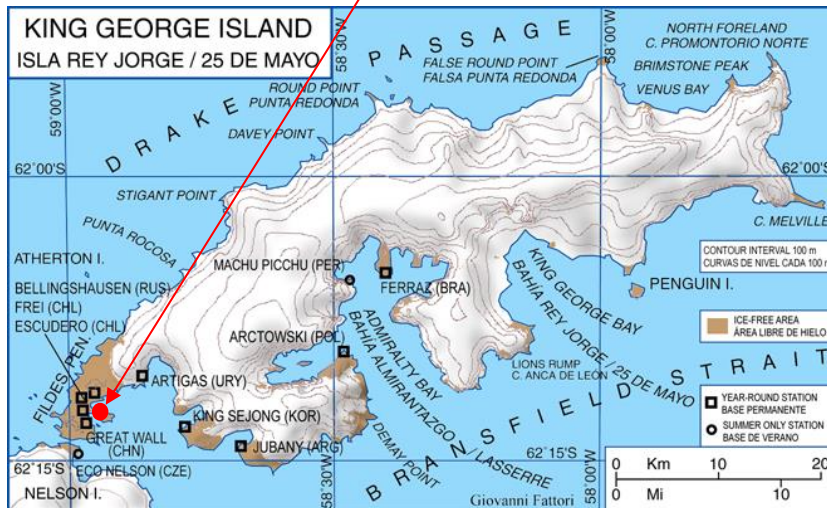


Fildes Bay, King George Island, Antarctica





# HAB species range expansion?

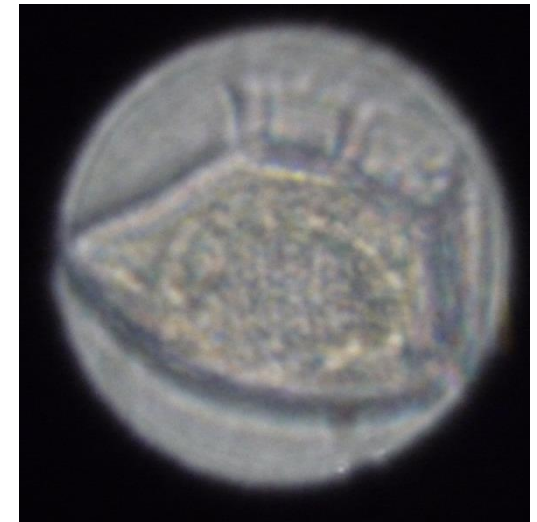




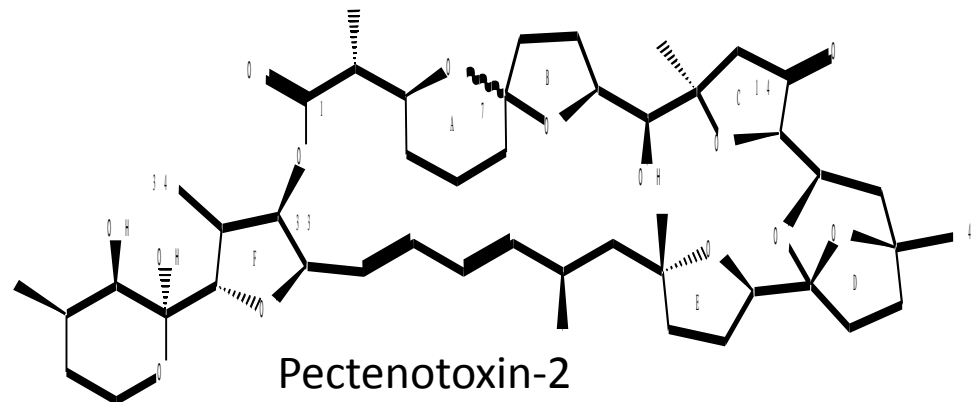
# HAB species range expansion?



## Plankton Net Haul



*Dinophysis norvegica*



*Dinophysis norvegica* recently has been found in the SW Atlantic  
Fabro et al. 2016. Harmful Algae 59,31-41.

- Gill damage
- O<sub>2</sub> deficiency
- secondary infections
- **Ichthyotoxins**



# Ichthyotoxic HAB species

## Raphidophyceans/Dictyochophyceans:

### *Chattonella*

*antiqua*

Ichthyotoxins

*globosa*

Ichthyotoxins

*marina*

Ichthyotoxins

*subsalsa*

Ichthyotoxins

*verruculosa*

Ichthyotoxins

### *Fibrocapsa*

*japonica*

Ichthyotoxins

### *Heterosigma*

*akashiwo*

Ichthyotoxins

### *Pseudochattonella*

c.f. *veruculosa*

Ichthyotoxins



*Chattonella antiqua*



*Fibrocapsa japonica*



*Heterosigma  
akashiwo*

**Toxins and exact mode of action unknown!**

# Ichthyotoxic HAB species

## Haptophytes:

*Chrysocromulina*

*leadbeateri*

*polylepis*

*Phaeocystis*

*pouchetii*

*Prymnesium*

*calathiferum*

*faveolatum*

*parvum*

*patelliferum*

*zebrinum*

Ichthyotoxins

Ichthyotoxins

Ichthyotoxins (?)

Ichthyotoxins

Ichthyotoxins

Ichthyotoxins, prymnesins (PRM)

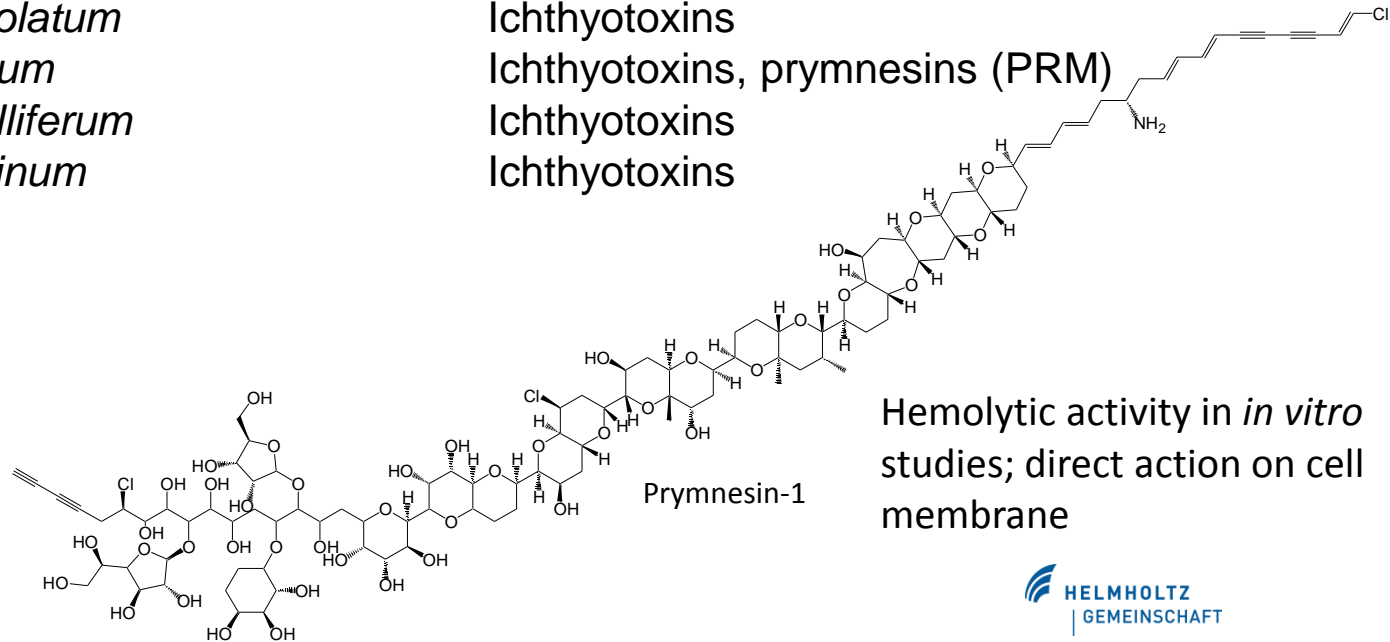
Ichthyotoxins

Ichthyotoxins

*Chrysocromulina  
polylepis*



*Prymnesium parvum*



Hemolytic activity in *in vitro*  
studies; direct action on cell  
membrane

**Toxins and exact mode of action unknown!**





## Thecate dinoflagellates:

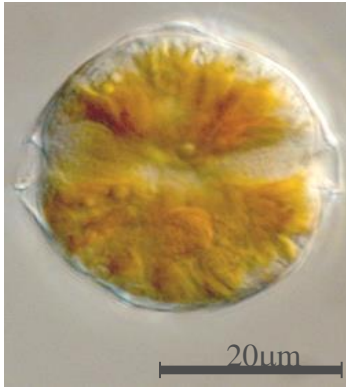
*Alexandrium*  
*catenella*  
*ostenfeldii*  
*Protoceratium*  
*reticulatum*



*Alexandrium ostenfeldii*

## Ichthyotoxins ?

## Lytic Effect



*Alexandrium catenella*  
strain 2 (Alex2)



*Rhodomonas salina*



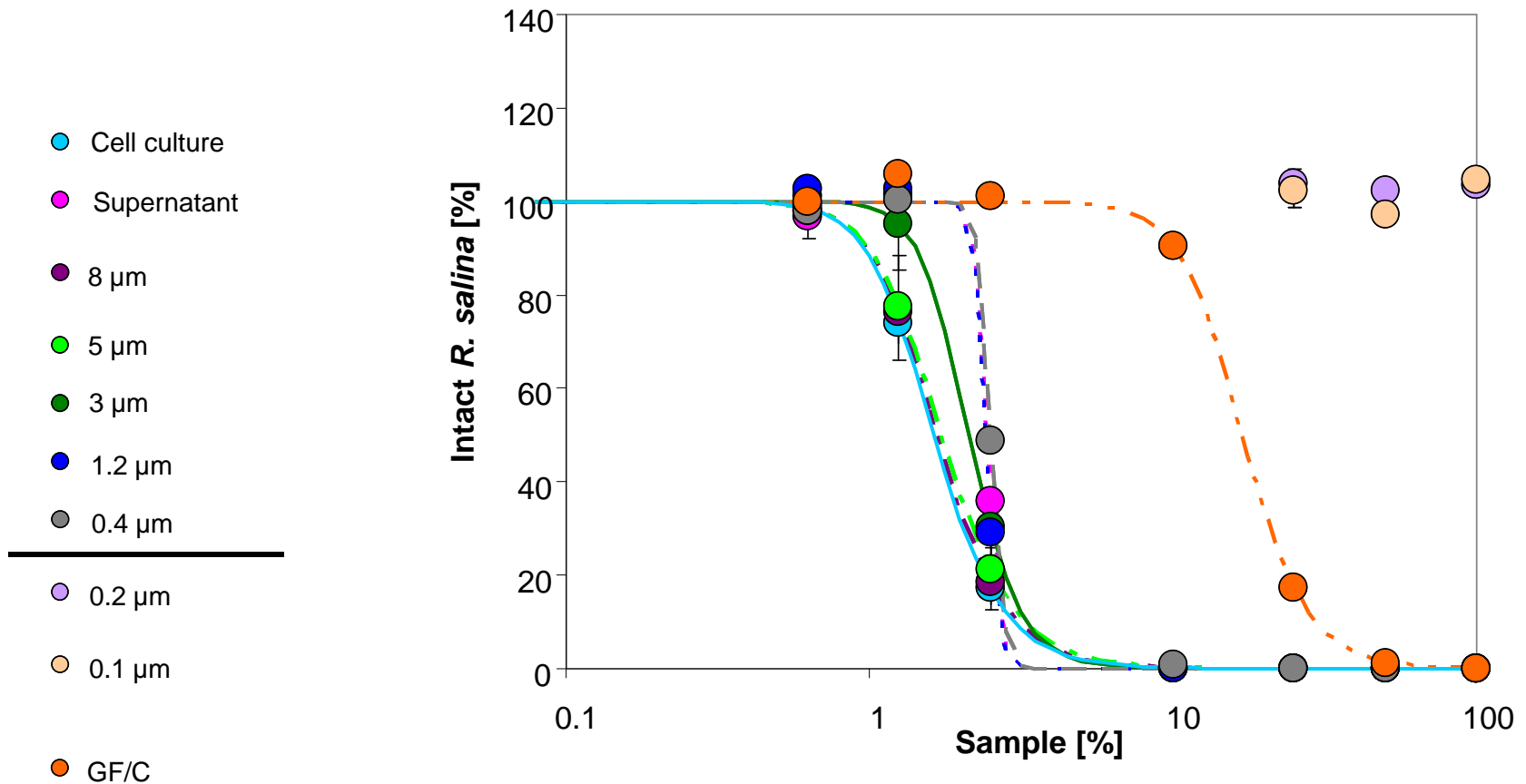
*Rhodomonas salina* exposed to *A. catenella* supernatant (cell free)



Video: U. Tillmann

## Characteristics

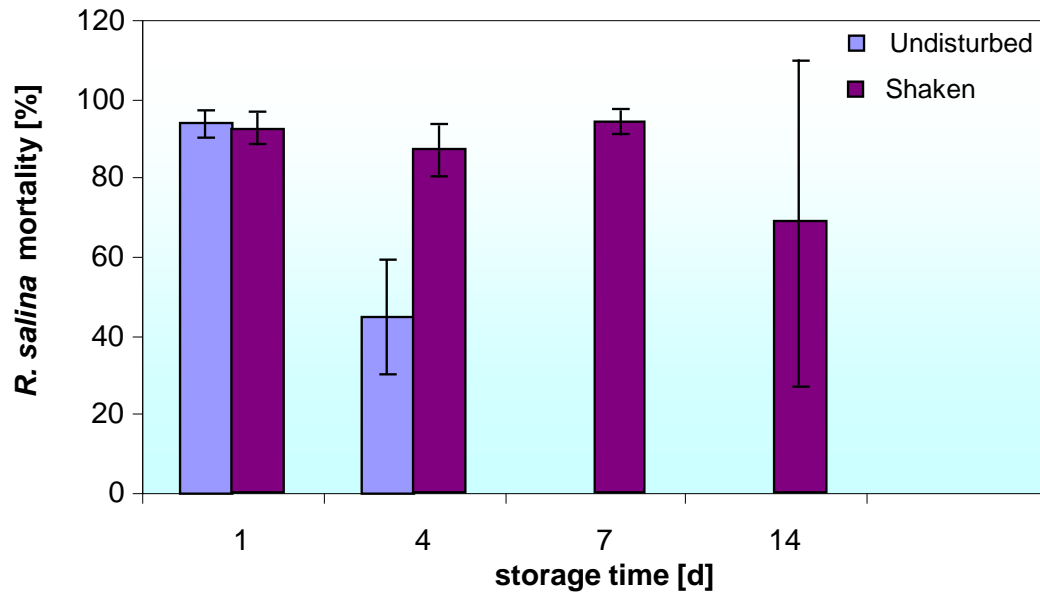
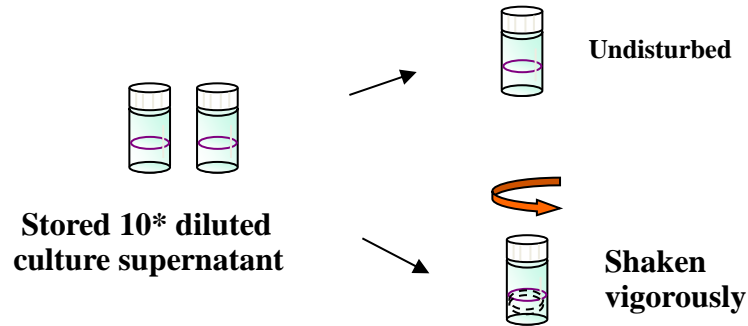
### Filtration





## Characteristics

Time



Lytic activity **cannot** be extracted by organic solvents from the aqueous supernatant,

but is enriched in a foamy emulsion between the aqueous and organic phases.

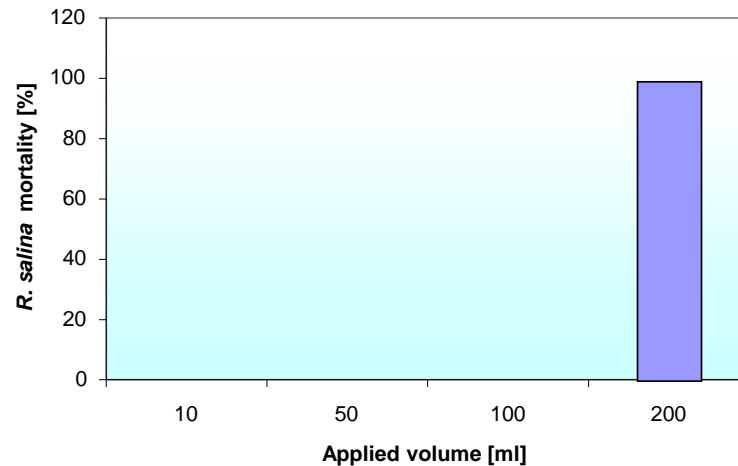
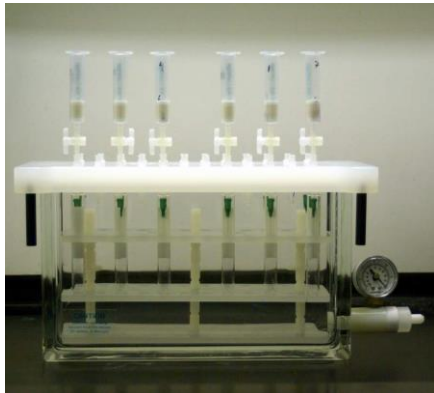
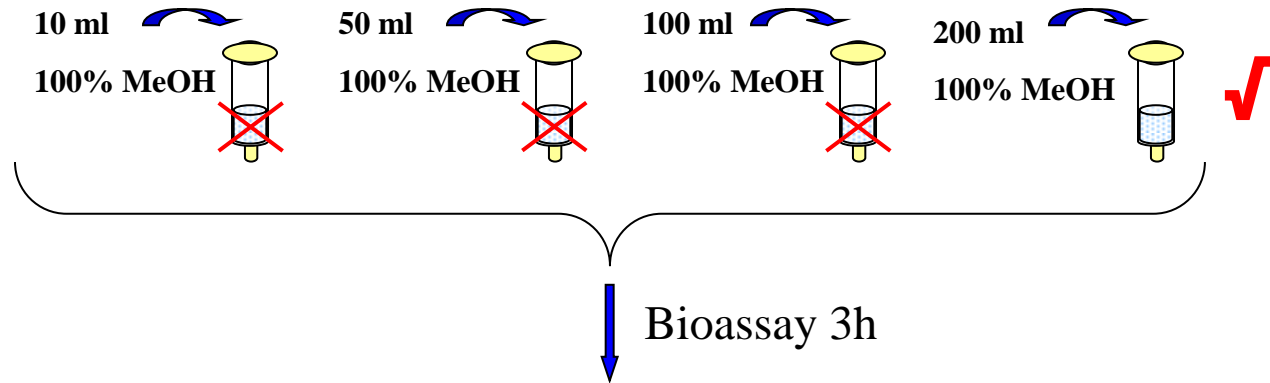
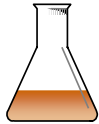
## Characteristics

- Liquid-liquid partitioning of supernatant (pH2) with n-hexane => no activity was extracted into the organic phase => **no lipids**
- Phenol sulfuric acid assay: saccharide content <2% => **no polysaccharides**
- Tryptic digestion did not change chromatographic behavior of lytic activity => **no proteins**

Ma et al. 2011. Harmful Algae 11, 65-72.

## Purification: Reversed phase SPE

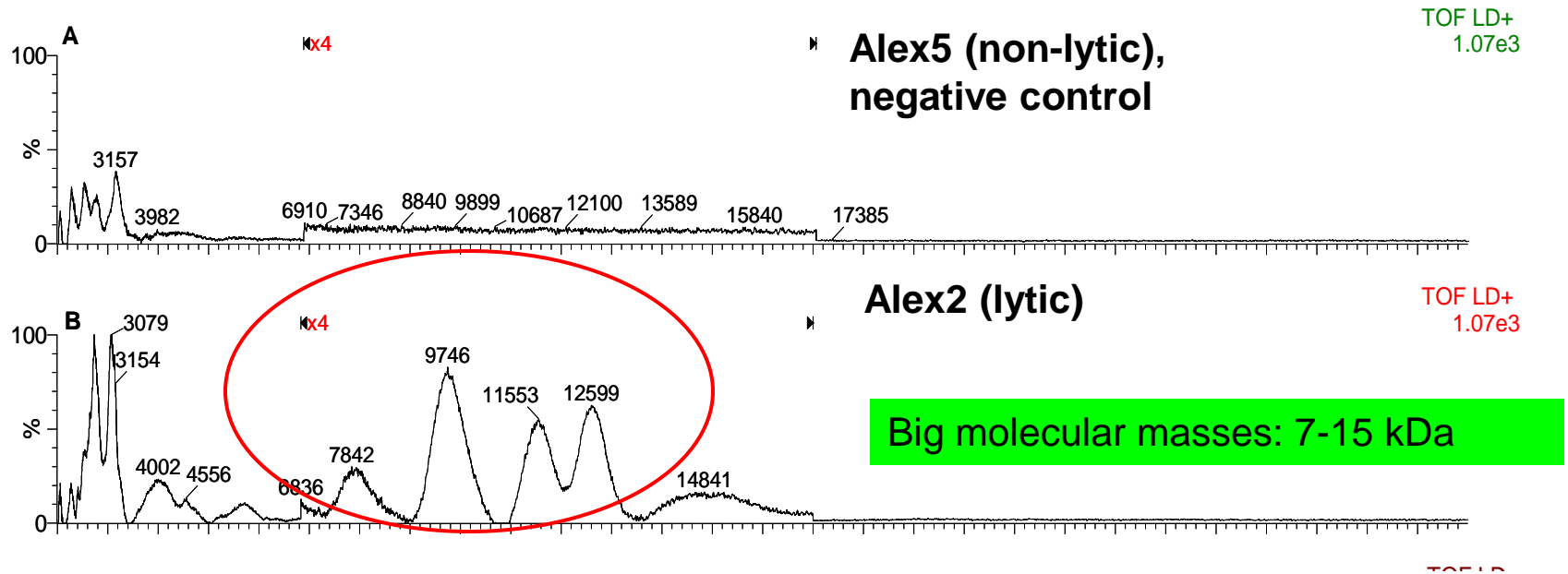
Alex2 supernatant



Allelochemicals „search“ their targets, i.e. specifically adsorb to lipophilic surfaces such as membranes

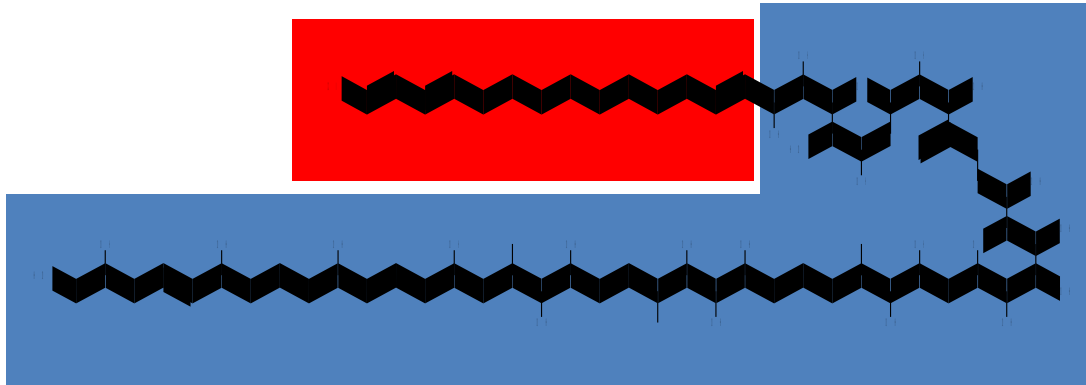
## Mass spectrometric characterization

### Time of Flight (TOF)-MS



MALDI-TOF mass spectra of HILIC fractions

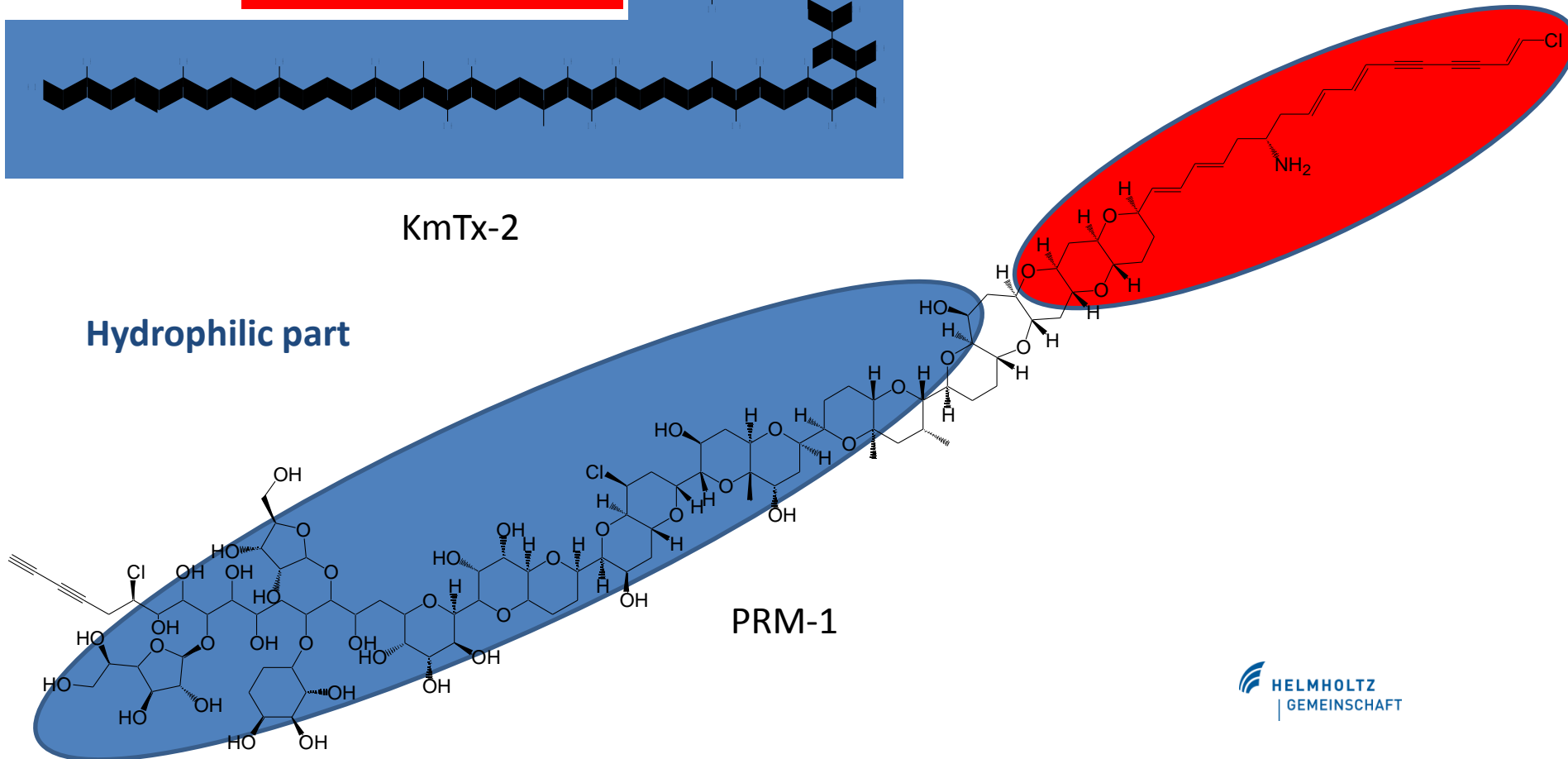
# Ichthyotoxic HAB species



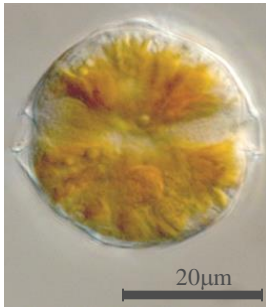
Lipophilic part

KmTx-2

Hydrophilic part



PRM-1







*Alexandrium*

- Paralytic shellfish poisoning (*Prakash et al., 1971*)
- Spirolides (*Cembella et al., 2000*).
- Gymnodimines (*Van Wagoner et al., 2011; Van de Waal et al. 2015*)
- Large scale fish kills (*Mortensen, 1985; Cembella et al., 2002*)
- Marine mammals mortality and morbidities (*Durbin et al., 2002; Doucette et al., 2006*)
- Allelopathy: toward other microalgae (*Blanco and Campos, 1988; Arzul et al., 1999; Fistarol et al., 2004; Tillmann et al., 2007*) and heterotrophic protists (*Hansen, 1989; Hansen et al., 1992; Matsuoka et al., 2000; Tillmann and John, 2002, Tillmann et al., 2007*).



# Take home messages

-  Toxin variability can be very high within a local population but also among populations of different geographic regions  
=> it is important to know local species and toxin variants
-  Plankton communities may rapidly change over time in a given region  
=> regular plankton monitoring including less abundant species is necessary
-  Is ichthyotoxicity a side effect of interplanktonic defense mechanisms?
-  Ichthyotoxins/allelochemicals need to be characterized to understand their fish killing activity and to develop mitigation strategies.

¡Gracias por su  
atención!

